Original Article

Dentistry Section

Comparative Evaluation of Different Antioxidants on Reversal of Microtensile Bond Strength of Composite Resin in Endodontically Treated Tooth Surface

RAMESH BHARTI¹, ANIL CHANDRA²

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ABSTRACT

Introduction: The use of Sodium Hypochlorite (NaOCI) as an endodontic irrigant changes the properties of dentine and lowers the bond strength of resin cements to the dentine, but it can be reversed by the use of antioxidants.

Aim: To evaluate the effect of antioxidants for restoring adhesion potential/reversal of microtensile bond strength of dentin in the pulp chamber treated with NaOCI and Ethylenediaminetetracetic Acid (EDTA).

Materials and Methods: This is a laboratory based experimental in-vitro study in which 40 freshly extracted human incisors were selected and cut to expose the pulp chamber below the dentine. The specimens were distributed among five groups: Group I samples were irrigated with 5.25% NaOCI for 30 minutes followed by 17% EDTA for 3 minutes and final rinse was done with 5.25% NaOCI (Positive control). Group II samples were treated with distilled water only (Negative control). In Group III, samples were treated same as in group I followed by 10%

INTRODUCTION

The goal of successful endodontic therapy is complete debridement of the root canal to remove inflamed and necrosed pulp tissue and achievement of apical as well as coronal hermetic seal [1]. Furthermore, immediately after the endodontic treatment coronal leakage is prevented by using permanent post-endodontics restoration, out of which restorative resin is most commonly used [2-4]. Endodontically treated tooth restored with composite resin showed better fracture resistance when compared to amalgam because of its ability to form a hybrid layer with dentine [5-7]. NaOCI is frequently used as a root canal irrigant because of its ability to dissolve the organic matter along with its antimicrobial property [8]. Nevertheless, this irrigant may affect the interaction of dentine to the adhesive restorative materials.

The bond strength of restorative resin decreases after using NaOCI due to release of residual oxygen which interferes with polymerisation of resin monomer that ultimately affect the dentine permeability, micro hardness and fracture resistance [9]. To minimise these adverse effects antioxidants can be applied before final restoration to neutralise and repeal of oxidising effect of NaOCI on dentine surface [10,11]. In dentistry Sodium Ascorbate (SA) is a commonly used antioxidant. SA improves the bond strength by neutralising and reversing the oxidising effect of NaOCI treated dentin [12].

Alpha tocopherol (vitamin E) an alternative of SA which is neutral, biocompatible and lipid soluble antioxidant. The ability of alpha tocopherol allowes polymerisation of the free radical and reverses the negative effect of NaOCI and improves resin bonding in endodontically treated teeth [13].

Sodium Ascorbate (SA) for 10 minutes. Samples of group IV were also treated as in group I followed by application of 5% solution of Alpha tocopherol for 10 minutes. Group V samples were treated same as in group I followed by a rinse with 5% solution of Sodium Thiosulphate ($Na_2S_2O_3$) for 10 minutes. Composite resin build up was performed and interface was tested by using Universal testing machine for Microtensile bond strength. The resulting data was analysed by one-way Analysis of Variance (ANOVA) and Mann-Whitney U test (p<0.05).

Results: The results demonstrated that irrigation with 5.25% NaOCI and 17% EDTA produce significant (p<0.05) reduction in resin dentin bond strength but this could be reversed by application of 5% $Na_2S_2O_3$. Highest bond strength was observed in group V (22.38±0.84) and lowest for group I (15.38±1.22).

Conclusion: Dentin bond strength was significantly reduced when bonding was performed immediately after use of NaOCI. Use of antioxidants after final irrigation with NaOCI can be recommended for better bonding of composite resin.

Keywords: Root canal, Sodium hypochlorite, Sodium thiosulfate

Recently, sodium thiosulfate has been used as an antioxidant to attain neutralisation of free radicals produced from NaOCI and also to restore the bond strength of composite resin to dentine [14]. It has some capability to neutralise the oxidising effect of free radicals on the applied surface by redox reaction thus, assisting in complete polymerisation of composite resin. Furthermore, it can react with oxidants to neutralise unpaired electrons and form a stable product. Sodium thiosulfate has been used in many microbiological and clinical studies to neutralise NaOCI indicating its biocompatibility. Thereby making it interesting to use it as an antioxidant to recover the bond strength lost because of endodontic irrigating materials [15].

There is evidence of effects of ascorbic acid solution on NaOCI and H_2O_2 treated dentin [16]. Limited research has been done to analyse the effect on bond strength to sodium thiosulfate treated root canal dentine [17]. Hence, the present study was conducted.

A null hypothesis was formulated stating that there is no difference in the bond strength to dentin treated with root canal irrigants and use of SA, alpha tocopherol and $Na_2S_2O_3$ would not affect the bond strength to dentin after root canal irrigation with NaOCI.

MATERIALS AND METHODS

This in-vitro study was done in the Department of Conservative Dentistry and Endodontics, King George's Medical University Lucknow, Uttar Pradesh, India from October 2018 to March 2019 and bond strength was tested at Central Institute of Plastics Engineering and Technology (CIPET) Lucknow. After taking the ethical approval from the Institutional Ethical Committee (Ref.no. 77th ECMII-B IMR faculty/p16) study was carried out on extracted human teeth.

Sample size calculation: The sample size was calculated by an online statistical calculator Dhand NK and Khatkar MS (2014) (http:// statulator.com/SampleSize/ss2P.html) for comparing two independent mean and the power of the study was 80% [18].

Forty extracted human maxillary incisors were taken for this study that were extracted for periodontal and prosthodontic reasons and were stored in 0.2% thymol and used within 6 months. The crowns of teeth were separated from the root at cemento-enamel junction using a diamond disc. The crowns were cut into two halves mesio-distally. The pulp tissue was removed and dentin surface was flattened with 180-grit and 600-grit silicon Carbide (3M Imperial) paper under running water for 30 seconds to standardise the smear layer of the dentin surface.

Samples were then divided into five groups according to the antioxidant used by simple randomisation method. (n=8). In negative control group no treatment with NaOCI (Vishal Dento care Pvt., Ltd.,) or EDTA (Meta Biomed) was done, rest all samples were treated with 5 ml of 5.25% of NaOCI for 30 minutes according to the protocol described by Correa ACP et al., and then immersed in the 17% EDTA for three minutes following which final rinse was performed with 5% NaOCI for one minute [15]. In Positive control group composite resin was bonded to NaOCI and EDTA treated tooth surface immediately without using any antioxidant. For antioxidant treatment Na₂S₂O₂ (Thermo fisher scientific India Pvt., Powai, Mumbai) group specimens were immersed in the 5 mL of 5% solution of Na₂S₂O₂. For α tocopherol (sigma- Aldrich, Bangalore, India) group, α tocopherol group was dissolved in the ethanol to prepare 5% of solution of alpha tocopherol and SA (Rolex chemical industries, Mumbai, India) was dissolved in the water to produce the 5% of the solution of SA. Samples were immersed in solution for 10 minutes according to groups divided [Table/Fig-1].

Groups	Irrigation protocol	Antioxidant procedure		
I	5.25% NaOCI for 30 minutes			
	+ 17% EDTA for 3 minutes	No antioxidant		
	+ 5.25% NaOCI for 1 minute			
П	No irrigant	No antioxidant		
Ш	Same as in Group I	5 mL of 5% solution of SA		
IV	Same as in Group I	5 mL of 5% solution of tocopherol		
V	Same as in Group I 5 mL of 5% solution of Na ₂ S ₂ O ₃			
[Table/Fig-1]: Experimental groups according to irrigation protocol and antioxidant used.				

Group I (Positive control): Dentin surface was irrigated with 5 mL of 5.25% NaOCI for 30 minutes and then immersed in 17% EDTA for three minutes then final rinse done with 5.25% NaOCI for one minute.

Group II (Negative control): No irrigation and no antioxidant was used.

Group III: After treatment as in group 1, freshly prepared 5 mL of 5% SA (Rolex chemical industries, Mumbai, India) was applied to the dentin surface for 10 minute [16].

Group IV: After treatment as in group I, 5 mL of 5% solution of Alpha Tocopherol (Sigm-aldrich, Bangalore, India) was applied to the dentin surface for 10 minutes.

Group V: After treatment as in group I, 5 mL of 5% solution of $Na_2S_2O_3$ (Thermo fisher scientific India Pvt., Powai, Mumbai) was applied to the dentin surface for 10 minute.

Bonding procedure: After pre-treatment with antioxidants, surfaces of all the specimens were dried with absorbent papers, then treated sections from each group were mounted on the acrylic block with dentine surface facing upwards [Table/Fig-2]. Tooth Surface was etched with 37% of phosphoric acid (Dentsply Caulk, Int. Milford DE, USA) for 20 seconds and bonding agent (Te-Econom Bond, Ivoclar

Vivadent) was applied on the surface for 10 seconds and light cured for 20 seconds. Ryle's tube of 18 gauge is cut to produce segments of 3 mm length and blocks of composite resins (Te-Econom plus Ivoclar Vivadent) were prepared over dentin surface and each one was light cured for 20 seconds with curing light (Ivoclar Vivadent Bluephase N). The blocks were stored in distilled water at 37°C for 24 hours before testing [13].



Bond strength measurement: Microtensile bond strength was measured by subjecting the specimens to the universal testing machine (Instron 3340, Instron Co., Canton, MA, USA) operated at the crosshead speed of 1 mm/minute and 5.00 N (Newton) load was applied until failure had occurred. Experiment for the measurement of Bond strength was done by author under supervision of Scientist of CIPET. It was the machine based reading so no issues of inter-examiner reliability occurred.

STATISTICAL ANALYSIS

The data was evaluated using Statistical Package for Social Sciences (SPSS) Software Version (19.0). Under the normal distribution, the data was analysed by one-way ANOVA (p=0.05). Comparison of Microtensile bond strength between the groups was explained by Mann-Whitney U test.

RESULTS

Statistical analysis revealed that there was a significant difference between the groups (p<0.05) thus null hypothesis was rejected. The mean Microtensile bond strength of different groups are summarised in [Table/Fig-3]. The positive control (Group I) exhibited the lowest mean bond strength value (15.38±1.22 MPa), closely followed by the group IV (17.90±1.54MPa) and highest mean value by the Na₂S₂O₃ Group V (22.38±0.84).

Groups	Mean±SD (Mpa)			
1	15.38±1.22			
Ш	21.49±1.48			
Ш	19.36±1.39			
IV	17.90±1.54			
V	22.38±0.84			
[Table/Fig-3]: Mean and standard deviation of micro-tensile bond strength of different groups (MPa). SD: Standard deviation				

The Mann-Whitney U test revealed that p value in Groups I vs II, I vs III, I vs IV and I vs V was statistically significant at p < 0.05. However, it was not significant between Groups III vs IV and Group II vs V [Table/Fig-4,5]. Intergroup comparison of mean microtensile bond strength between Group III, IV and V is shown in [Table/Fig-5].

Groups	Z	p-value			
l vs ll	3.31	0.00094			
I vs III	3.31	0.00094			
I vs IV	2.47	0.01352			
I vs V	3.31	0.00094			
[Table/Fig-4]: Comparison of Microtensile bond strength in different groups using					

(p<0.05 is statistically significant, p<0.001 is statistically highly significant)

Groups	Z	p-value			
II vs V	1.103	0.27134			
III vs IV	1.418	0.1556			
III vs V	3.20	0.00138			
IV vs V	3.31	0.00094			
[Table/Fig-5]: Intergroup comparison of Microtensile bond strength in different groups using Mann-Whitney U Test.					

DISCUSSION

This study is one of the few studies that evaluate the resin bond strength to dentin and pulp chamber after the use of common endodontic irrigants, and it is the first to assess the comparison of three different antioxidant like sodium thiosulfate, SA, and alpha tocopherol to restore the lost dentin bond strength. The methodology was adjusted to compare the effect of different antioxidants on the area of the tooth surface which is fully affected by chemical substance during endodontic treatment [19].

Previous researches [20-24] as in [Table/Fig-6] have shown the detrimental effect of different root canal irrigants on the bond strength of postendodontic restorative resin cement to the radicular/coronal dentin. In a study done by Erdemir A et al., they found that 3% hydrogen peroxide significantly lowered the bond strength [25].

of the treated substrate and facilitating the complete polymerisation of adhesive material [28]. On the other hand it can react with antioxidants to neutralise unpaired electrons and form a stable product [29]. Then, it becomes important to use this antioxidant substance to recover the bond strength lost due to the use of endodontic irrigants. Na₂S₂O₂ can be used up to 6 months when stored in a refrigerator [30].

SA is capable of reducing a variety of oxidative compounds, particularly free radicals [16]. It is used just immediately after preparation because of its short shelf life. The antioxidant ability of SA can help to neutralise and reverse the oxidising property of NaOCI. In the present study, SA allows free-radical polymerisation of resins to proceed without premature termination hence reverse the action of NaOCI [10].

In case of bleached enamel, Suneetha R et al., studied 10% SA solution as an antioxidant for restoring the shear bond strength immediately after bleaching [31]. Subramanian R et al., observed that after application of 10% SA just after beaching could neutralize the residual oxygen and restore the bond strength [32]. In another study it was also observed that in scanning electron microscopic images ascorbic acid causes excessive etching of bleached enamel surface [12].

Vitamin E is the term used for a group of tocopherols and tocotrienols, of which α tocopherol has the highest biological activity. Vitamin E functions as a chain-breaking antioxidant that prevents propagation of free radical reactions [33]. Vitamin E (alpha-tocopherol), same as Ascorbic acid restored the bond strength by removing the free radicles molecular oxygen. It has recently been suggested for improving composite bonding with coronal dentine after endodontic treatment [33].

Results of this study showed that group V and group II (Negative Control) have higher bond strength followed by group III and

Author and Year	Test modality	Antioxidants agent used	Application time	Concentration of NaOCI/ EDTA used	Results		
Lai SC et al., 2001[9]	Microtensile bond strength	10% sodium ascorbate solution	10 minutes	5% NaOCI	Effective		
Soeno K et al., 2004 [19]	Tensile bond strength	10% ascorbic acid	15-60 seconds	10% NaOCI	Effective		
Weston CH et al., 2007 [11]	Tensile bond strength	10% ascorbic acid	10 minutes	5.25% NaOCI	Effective		
da Cunha LF et al., 2010 [21]	Push out bond strength	10% ascorbic Acid	10 minutes	5% NaOCI	Effective		
Manimaran VS et al., 2011[24]	Microtensile bond strength	10% sodium ascorbate	10 minutes	5.25% NaOCI	Effective		
Prasansuttiporn T et al., 2011 [27]	Microtensile bond Strength	10% sodium ascorbate Solution/ Rosmarinic acid solution	5-10 minutes	6% NaOCI	Effective		
Khoroushi M and Kachuei M [20]	Tensile bond strength	10% sodium ascorbate Hydrogel	2 minutes	2.5% NaOCI	Effective		
Stevens CD 2014 [22]	Shear Bond Strength	10% sodium ascorbate	1 minutes	6% NaOCI	Effective		
Gonulol N et al., 2015 [16]	Microtensile Bond strength	10% sodium ascorbate solution	10 minutes	5.25% NaOCI	Effective		
Ebrahimi-Chaharom ME et al., 2015 [23]	Shear bond strength	10% sodium ascorbate	10 minutes	5.25% NaOCI	Effective		
Correa ACP et al., 2016 [15]	Microtensile bond strength	0.5% or 5% sodium thiosulfate	1, 5 or 10 minutes	5.25% NaOCI	Effective		
Bharti R and Chandra A (Current Study)	Microtensile bond strength	5% solution of sodium ascorbate, sodium thiosulfate and Alpha tocopherol	10 minutes	5.25% NaOCI	Effective		
[Table/Fig-6]: Description of in-vitro studies using different antioxidants/reducing agents to revert the decrease in bond strength after use of NaOCI/EDTA in root canal wall dentin [9,11,15,16,19-24,27].							

Use of NaOCI on the surface of coronal dentine causes changes in the mechanical properties of dentin, such as micro hardness, elastic modulus, and fracture resistance that might produce a "new" dentin surface and this may be due to damage caused to the organic matrix, collagen, and finally leaving mineralised coronal dentine surface [26]. Decreased bond strength is the result of the oxidising action of NaOCI. It may be possible for the compromised bond strength to be reversed by a reduction of the oxidised surfaces with an antioxidant such as Na₂S₂O₃, SA, and alpha tocopherol [27].

Na₂S₂O₂ has been used in many microbiology studies to neutralise NaOCI and it is a potent antioxidant. It is possible that it has the potential to neutralise the oxidising agents through redox reaction

Group IV. SA also reversed the bond strength in this study which is supportive to the previous study [34]. Alfa tocopherol was used as a gel in this study. It also resulted in the bond strength reversal but less than in group III (SA). This may be due to the nonaqueous nature of this antioxidant.

Limitation(s)

This study did not evaluate the bond strength before the use of antioxidants. As this is an invitro study, clinical evaluation is necessary to determine the efficacy of different antioxidants in restoring the microtensile bond strength. Sodium ascorbate has a short shelf-life so it can not be stored for a long time, it should be prepared freshly whenever it is to be used for better results.

CONCLUSION(S)

The reversal effects of $Na_2S_2O_3$, ascorbic acid and alpha tocopherol on the compromised bond strength in NaOCI/EDTA-treated dentin using the total etching adhesive system were found to be satisfactory. Nevertheless, the use of 5% $Na_2S_2O_3$ for 10 minutes showed the best result compared to ascorbic acid and alpha tocopherol. These findings are clinically important/significant because they suggest SA, alpha tocopherol and $Na_2S_2O_3$ can be used for reversal of bond strength of post-endodontic restorative resin to root canal treated dentin surface. Considering the available literature and the limitations of each antioxidants under consideration, none of them completely reverse the bond strength to its original. So, development of new antioxidants will help in safe and effective use before the postendodontic restoration.

REFERENCES

- Moreno JO, Alves FR, Gonçalves LS, Martinez AM, Rôças IN, Siqueira JF Jr. Periradicular status and quality of root canal fillings and coronal restorations in an urban Colombian population. J Endod. 2013;39(5):600-04.
- [2] Belli S, Zhang Y, Pereira PN, Pashley DH. Adhesive sealing of the pulp chamber. J Endod. 2001;27(8):521-26.
- [3] Belli S, Zhang Y, Pereira PN, Ozer F, Pashley DH. Regional bond strengths of adhesive resins to pulp chamber dentin. J Endod. 2001;27(8):527-32.
- [4] Galvan RR Jr, West LA, Liewehr FR, Pashley DH. Coronal microleakage of five materials used to create an intracoronal seal in endodontically treated teeth. J Endod. 2002;28(2):59-61.
- [5] Ausiello P, De Gee AJ, Rengo S, Davidson CL. Fracture resistance of endodontically-treated premolars adhesively restored. Am J Dent. 1997;10(5):237-41.
- [6] Schwartz RS. Adhesive dentistry and endodontics. Part 2: Bonding in the root canal System–The promise and the problems: A review. J Endod. 2006;32(12):1125-34.
- [7] Trope M, Langer I, Maltz D, Tronstad L. Resistance to fracture of restored endodontically treated premolars. Endod Dent Traumatol. 1986;2(1):35-38.
- [8] Zehnder M. Root canal irrigants. J Endod. 2006;32(5):389-98.
- [9] Lai SC, Mak YF, Cheung GS, Osorio R, Toledano M, Carvalho RM, et al. Reversal of compromised bonding to oxidizoxidised etched dentin. J Dent Res. 2001;80(10):1919-24.
- [10] Vongphan N, Senawongse P, Somsiri W, Harnirattisai C. Effects of sodium ascorbate on microtensile bond strength of total-etching adhesive system to NaOCI treated dentine. J Dent. 2005;33(8):689-95.
- [11] Weston CH, Ito S, Wadgaonkar B, Pashley DH. Effects of time and concentration of sodium ascorbate on reversal of NaOCI-induced reduction in bond strengths. J Endod. 2007;33(7):879-81.
- [12] Muraguchi K, Shigenobu S, Suzuki S, Tanaka T. Improvement of bonding to bleached bovine tooth surfaces by ascorbic acid treatment. Dent Mater J. 2007;26(6):875-81.
- [13] Kavitha M, Selvaraj S, Khetarpal A, Raj A, Pasupathy S, Shekar S. Comparative evaluation of superoxide dismutase, alpha-tocopherol, and 10% sodium ascorbate on reversal of shear bond strength of bleached enamel: An in vitro study. Eur J Dent. 2016;10(1):109-115.
- [14] Gomes BP, Martinho FC, Vianna ME. Comparison of 2.5% sodium hypochlorite and 2% chlorhexidine gel on oral bacterial lipopolysaccharide reduction from primarily infected root canals. J Endod. 2009;35(10):1350-53.

- [15] Corrêa ACP, Cecchin D, de Almeida JFA, de Almeida Gomes BPF, Zaia AA, Ferraz CCR. Sodium thiosulfate for recovery of bond strength to dentin treated with sodium hypochlorite. J Endod. 2016;42(2):284-88.
- [16] Gonulol N, Kalyoncuoglu E, Ertas E. Effect of sodium ascorbate on dentin bond strength after treatment with oxidising root canal irrigants. J Dent Sci. 2015;10:139-44.
- [17] Gharge AR, Mattigatti S, Mangala TM. In-vitro comparative evaluation of reducing agents for recovery of bond strength to dentin treated with sodium hypochlorite. International Journal of Science and Research (IJSR). 2018;7(5):1473-76. https:// www.ijsr.net/search_index_results_paperid.php?id=ART20182651.
- [18] Dhand NK, Khatkar MS. Statulator: An online statistical calculator. Sample size calculator for comparing two independent proportions. 2014 http://statulator. com/SampleSize/ss2P.html.
- [19] Soeno K, Taira Y, Matsumura H, Atsuta M, Suzuki S. Adhesion of 4-META/MMA-TBB resin to collagen-depleted dentin-Effect of conditioner with ascorbic acid/ ferric chloride. Dent Mater J. 2004;23(2):100-05.
- [20] Khoroushi M, Kachuei M. Pull-out bond strength of a self-adhesive resin cement to NaOCI-treated root dentin: Effect of antioxidizoxidising agents. Restor Dent Endod. 2014;39(2):95-103.
- [21] da Cunha LF, Furuse AY, Mondelli RF, Mondelli J. Compromised bond strength after root dentin deproteinization reversed with ascorbic acid. J Endod. 2010;36(1):130-34.
- [22] Stevens CD. Immediate shear bond strength of resin cements to sodium hypochlorite-treated dentin. J Endod. 2014;40(9)1459-62.
- [23] Ebrahimi-Chaharom ME, Kimyai S, Mohammadi N, Oskoee PA, Dane-shpuy M, Bahari M. Effect of sodium ascorbate on the bond strength of all-in-one adhesive systems to NaOCI-treated dentin. J Clin Exp Dent. 2015;7(5):e595-99.
- [24] Manimaran VS, Srinivasulu S, Rajesh Ebenezar AV, Mahalaxmi S, Srinivasan N. Application of a proanthocyanidin agent to improve the bond strength of root dentin treated with sodium hypochlorite. J Conserv Dent. 2011;14(3):306-08.
- [25] Erdemir A, Ari H, Güngüneş H, Belli S. Effect of medications for root canal treatment on bonding to root canal dentin. J Endod. 2004;30(2):113-16.
- [26] Moreira DM, Almeida JF, Ferraz CC, Gomes BP, Line SR, Zaia AA. Structural analysis of bovine root dentin after use of different endodontics auxiliary chemical substances. J Endod. 2009;35:1023-27.
- [27] Prasansuttiporn T, Nakajima M, Kunawarote S, Foxton RM, Tagami J. Effect of reducing agents on bond strength to NaOCI-treated dentin. Dent Mater. 2011;27(3):229-34.
- [28] Morris MD, Lee KW, Agee KA, Bouillaguet S, Pashley DH. Effects of sodium hypochlorite and RC-prep on bond strengths of resin cement to endodontic surfaces. J Endod. 2001;27(12):753-57.
- [29] Aruoma OI, Cuppett SL. Antioxidant Methodology: In Vivo and In-vitro Concepts. Champaign, IL: AOCS Press; 1997.
- [30] Sooriyaarachchi M, Narendran A, Gailer J. The effect of sodium thiosulfate on the metabolism of cis-platin in human plasma in-vitro. Metallomics. 2012;24(9):960-67.
- [31] Suneetha R, Pavithra S, Thomas J, Nanga GS, Shiromany A, Shivrayan A. An In Vitroin-vitro comparative study of shear bond strength of Ccomposite resin to bleached enamel using synthetic and herbal antioxidants. J Int Oral Health. 2014;6(6):77-81.
- [32] Subramonian R, Mathai V, Christaine Angelo JB, Ravi J. Effect of three different antioxidants on the shear bond strength of composite resin to bleached enamel: An in-vitro study. J Conserv Dent. 2015;18(2):144-48.
- [33] Brigelius-Flohe R, Traber MG. Vitamin E: function and metabolism. FASEB J. 1999;13(10):1145-55.
- [34] Lai SC, Tay FR, Cheung GS, Mak YF, Carvalho RM, Wei SH, et al. Reversal of compromised bonding in bleached enamel. J Dent Res. 2002;81(7):477-81.

PARTICULARS OF CONTRIBUTORS:

- 1. Additional Professor, Department of Conservative Dentistry and Endodontics, Faculty of Dental Sciences, King George's Medical University, Lucknow, Uttar Pradesh, India.
- 2. Professor, Department of Conservative Dentistry and Endodontics, Faculty of Dental Sciences, King George's Medical University, Lucknow, Uttar Pradesh, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Ramesh Bharti,

Additional Professor, Department of Conservative Dentistry and Endodontics,

Faculty of Dental Sciences, King George's Medical University, Lucknow, Uttar Pradesh, India.

E-mail: rameshbharti@kgmcindia.edu

AUTHOR DECLARATION:

- Financial or Other Competing Interests: None
- Was Ethics Committee Approval obtained for this study? Yes
- Was informed consent obtained from the subjects involved in the study? NA
- For any images presented appropriate consent has been obtained from the subjects. NA

PLAGIARISM CHECKING METHODS: [Jain H et al.]

- Plagiarism X-checker: Sep 16, 2020
- Manual Googling: Jan 27, 2021
- iThenticate Software: Apr 24, 2021 (20%)

Date of Submission: Sep 14, 2020 Date of Peer Review: Oct 23, 2020 Date of Acceptance: Mar 17, 2021 Date of Publishing: May 01, 2021

ETYMOLOGY: Author Origin